







- EU SPIDIA Project Update -

Standardization and Improvement of Generic Preanalytical Tools and Procedures for In Vitro Diagnostics

5th Annual BRN Symposium Bethesda, February 22nd 2012

Dr. Uwe Oelmueller SPIDIA Coordinator (QIAGEN)

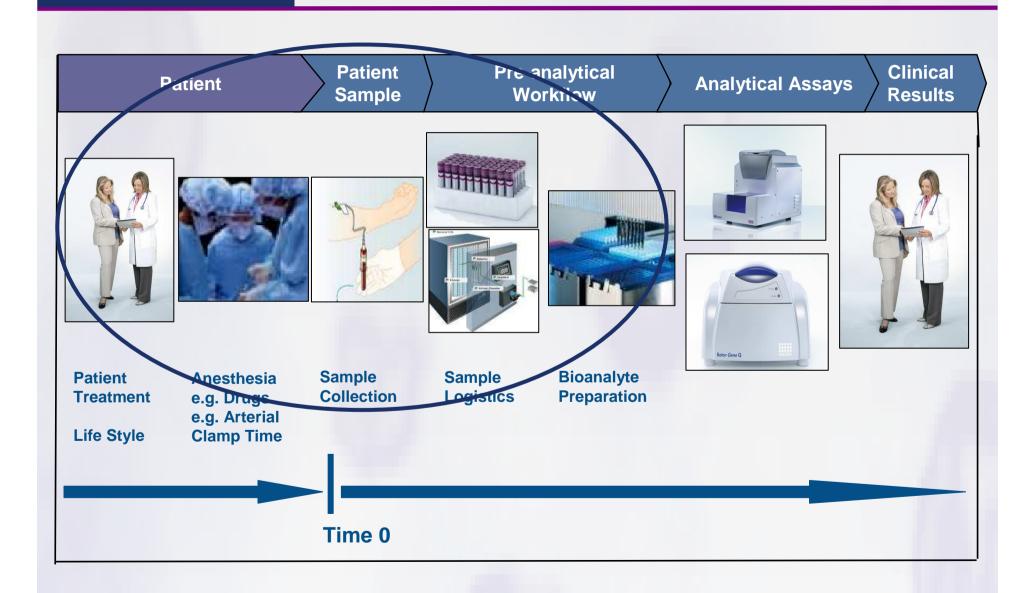


Agenda

- **SPIDIA Project History and Goals**
- Results & Status
 - New Technologies & Tools
 - Pan-European Guidelines
 - Biospecimen Quality Markers



Diagnostic WorkflowFrom Patients to Clinical Results

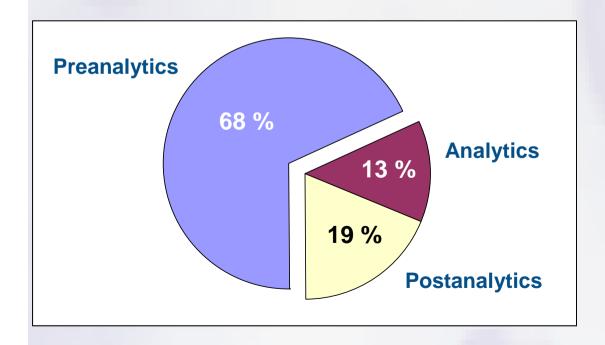




It is Real Problem

"Preanalytical errors still account for nearly 60%-70% of all problems occurring in laboratory diagnostics, most of them attributable to mishandling procedures during collection, handling, preparing or storing the specimens".

Lippi G. *et al.*. Preanalytical quality improvement: from dream to reality. Clin Chem Lab Med. 2011 Jul; 49(7):1113-26. Epub 2011 Apr 25.



Costs of ~ 460,000 \$ / year in an average German hospital caused by pre-analytical errors

Frost & Sullivan 2011 on behalf of BD



Project Main Goals

- Pan-European guidelines for preanalytics (Molecular Blood, Tissue)
- New pre-analytical tools & technologies (Blood, Plasma, Tissue, Swabs)
- Sample quality markers (Blood, Tissue)
- Training and dissemination



Project Facts

Program
European Commission FP7-HEALTH

Consortium 7 public research organizations

8 companies

1 standards organization (CEN)

Coordinator QIAGEN GmbH

Run Time
October 2008 – September 2012

(prolongation request intended)

■ Budget 13 Mio € (9 Mio € EC contribution)

Co-operations
NCI / OBBR, CLSI, EFCC, BBMRI and other

international initiatives and organizations

Web page <u>www.spidia.eu</u>

Newsletter



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New Tissue Fixation & Stabilization Histomorphology, IHC, RNA & DNA, Proteins

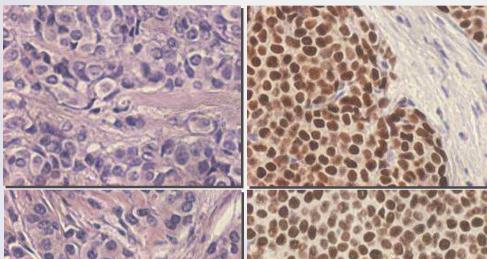
H&E Staining IDC of Breast

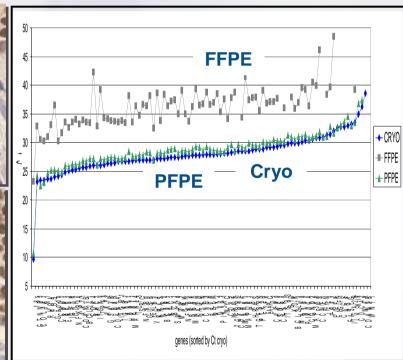
Estrogen Receptor a (clone 1D5) IDC of Breast

Mammacarcinoma
TaqMan Array Gene Signature

Formalin

AXgene





PFPE revealed preservation of morphology and antigenicity comparable to FFPE

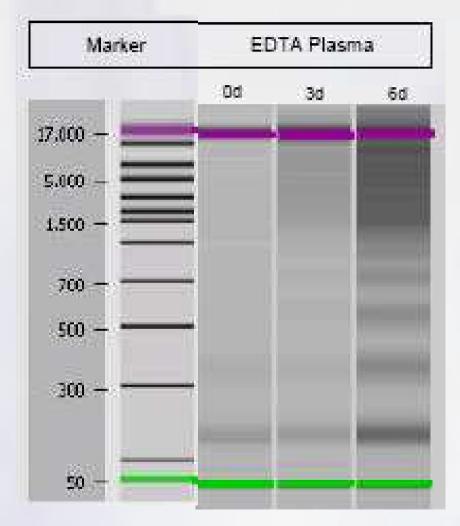
Nucleic acid analysis superior to FFPE

Kap M. et al., PLoS ONE 6(11): e27704 (2011) Viertler C. et al., submitted for publication

Groelz D. et al., unpublished data.



fcNA Profiles in Whole Blood / Plasma What is missing?



Horlitz M. et al., unpublished data

- Studies for understanding fcDNA and fcRNA profile stability / changes in whole blood and in plasma
- Development of fcDNA and fcRNA profile preservation technologies

EDTA blood was incubated for up to 6 days at room temperature. Blood fcDNA pattern stability was determined by separating the purified plasma DNA on a 2100 Agilent Bioanalyzer



Ongoing Technology & Tools Developments for Other Sample Types

■ Fine Needle Aspirates

Stabilization of morphology, antigenicity, DNA, RNA, proteome

Whole Blood

Stabilization of cell morphology and biomolecule profiles

Swabs

 Stabilization and improved processing of respiratory and samples for molecular analysis

Stabilized Whole Blood

Integrated automated sample-to-result workflows (cellular RNA, ncRNAs incl. miRNAs)



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Evidence Based Guidelines Examples Blood DNA & RNA, Plasma fcDNA

- Phase 1 Trials Laboratories used their workflows & tools
- Let by Prof. Pazzagli (Univ. Florence), supported by the EFCC



- Guidelines / Standards Concepts CEN
- Phase 2 Trials Laboratories will use SPIDIA's optimized workflows
- Guidelines / Standards Developments CEN

SPIDIA Trials	No. of Participants (29 countries)	Participants who sent NA samples back	Percentage of NA samples sent back
Blood RNA	102	93	91 %
Blood DNA	130	121	93 %
Plasma DNA	67	62	93 %
Total	299	276	92 %



Blood DNA Trial 1 - Examples for Pre-analytical Workflow Variations

■ Blood storage time before DNA extraction

• 39 labs: ≤ 6 days

• 60 labs: 6 – 10 days

• 53 labs: ≥ 10 days

■ Blood storage temperature before DNA extraction

• 18 labs: -20 °C

• 129 labs: +4 °C

• 9 labs: ambient temp.

■ Isolated DNA storage before analysis

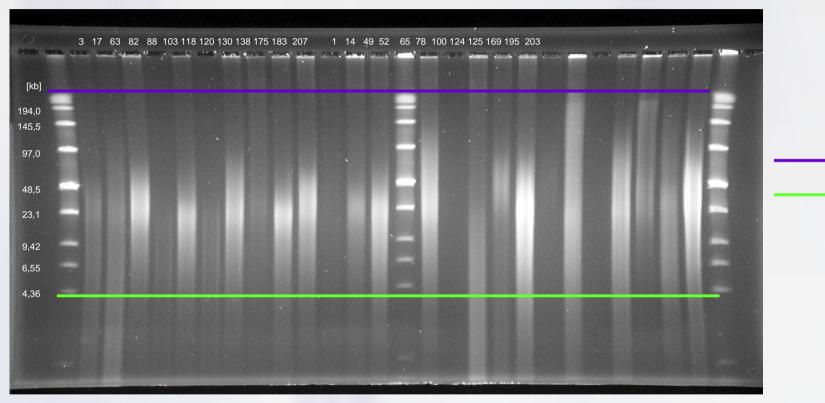
• 20 labs: -20 °C

• 111 labs: +4 °C

• 27 labs: ambient temp.



DNA Length Variation – Pulse Field Gel Electrophoresis



195 kb 4.36 kb

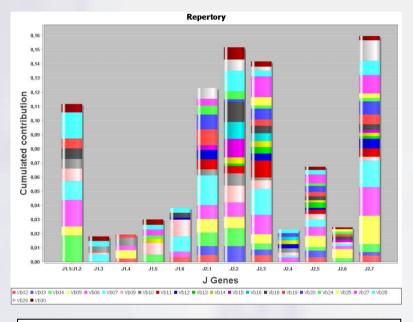
- High molecular weight DNA integrity: degradation, fragmentation
- High variability among samples

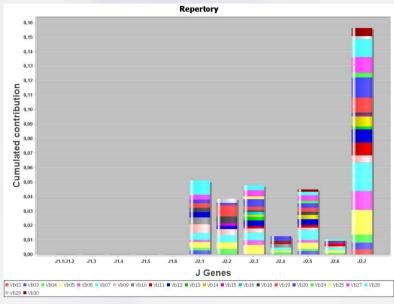
Hartmann C. *et al.*, unpublished results Pazzagli M. et al., manuscript in preparation



Impact of DNA quality on Immune T cell Repertoire Analysis (ImmunID Technologies)

V contribution for each J gene – Research Trial (ImmunID Technologies, France)





Ref. DNA from UNFI (DIV 54%)

Sample 38 (Poor quality) (DIV 32%)

- Lost of all long V—J rearrangements
- Lost of part of intermediate length rearrangements

L. Barraud et al. Unpublished data Pazzagli M. et al., manuscript in preparation



Blood RNA Ring Trial Parameters

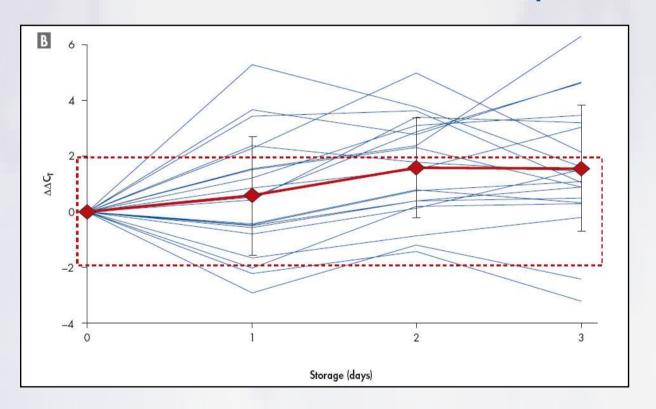
- Purity
- Interference substances (RT-qPCR)
- Yield
- Integrity
- RNA Profile Stability / Changes



Changes of Transcripts Profiles in Blood

Individual Samples React Differently

Human EDTA Blood stored at Room Temperature over 3 days



IL-1β mRNA

Guenther K. et al.. AMP Poster (2005)



Learning from Blood RNA Ring Trial 1

- No pooling of different donors' blood
 - Accept that only sub-groups of ring trial participating laboratories get the same blood samples
- No usual blood collection bags
 - Use dedicated EDTA bags
- Immediate cooling of blood bags
 - Artificial gene induction and down regulation to be avoided
- Use of intracellular RNA markers
 - External markers will behave differently



Blood RNA Second Ring Trial

Preparation of Blood Samples

1 bag - 1 donor





PAXgene Blood RNA Tubes



Empty bag

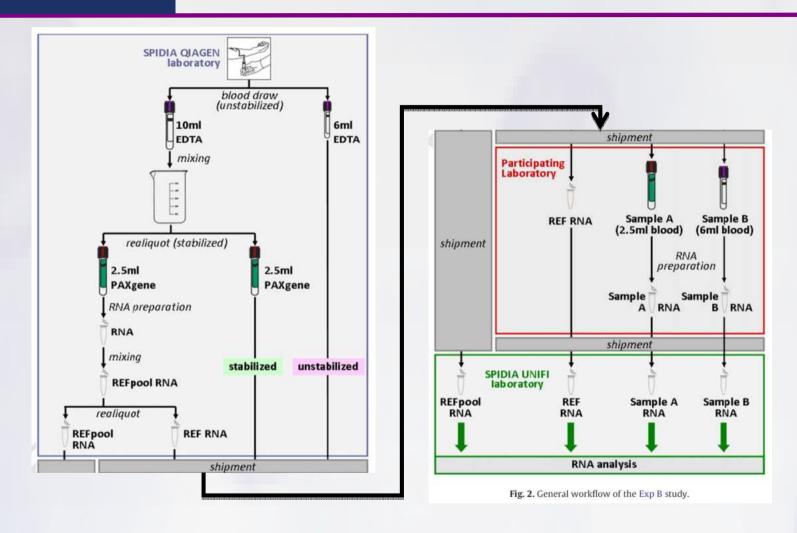
- Filled with 39 ml of EDTA solution under sterile condition
- Filled with 461 ml blood from phlebotomy







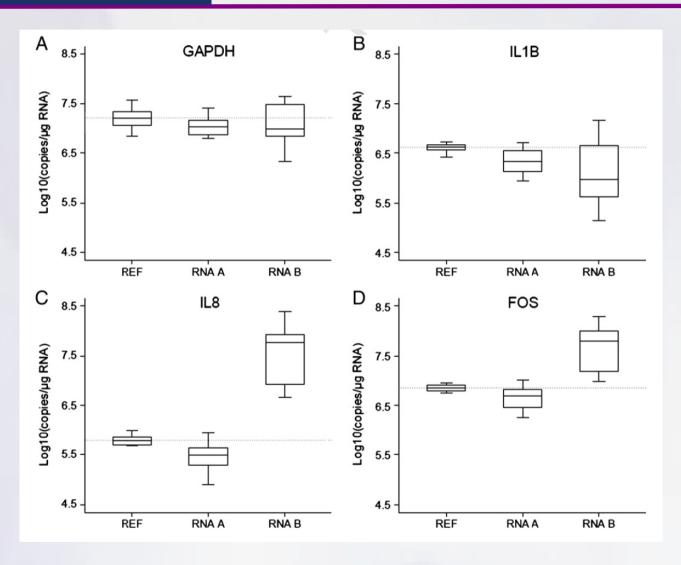
Proficiency Testing for Preanalytical Workflows used for Blood RNA Analysis



K. Günther, F. Malentacchi, P. Verderio, S. Pizzamiglio, C. M. Ciniselli, A. Tichopad, M. Kubista, R. Wyrich, M. Pazzagli, S. Gelmini. Implementation of a proficiency testing for the assessment of the preanalytical phase of blood samples used for RNA based analysis. Clin Chim Acta (2012) – in press.



Blood Sample Shipment - RNA Profile Changes Stabilized vs. EDTA Blood



Box plots reflecting the mRNA expression of GAPDH (Panel A), IL1B (Panel B), IL8 (Panel C), and FOS (Panel D) measured in the three sample types REF, RNA A (PAXgene Blood RNA) and RNA B (EDTA). Each box indicates the 25th and 75th percentiles. The horizontal line inside the box indicates. the median, and the whiskers indicate the extreme measured values. The dotted horizontal line indicates the median value of the REF samples (prior shipment) and serves for comparison.

Guenther K. et al. Clin Chim Acta. 2012, in press.



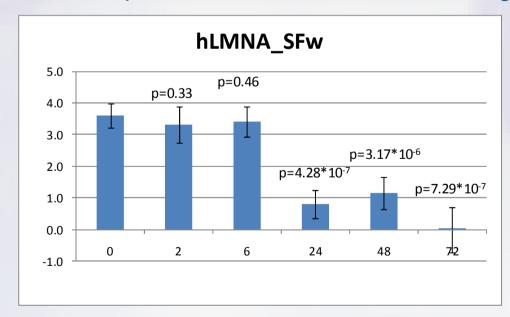
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Blood RNA Quality Marker Discovery

- Quality markers measuring RNA up- & down-regulation
 - >180 micro arrays (time course experiments)
 - 11 marker candidates (specific RNA degradation or gene down regulation, specific RNA gene induction, random degradation)
 - Technical assay validation
 - Next step: Performance validation within larger donor cohorts



Rian E. et al., unpublished data



Acknowledgement SPIDIA Consortium Members

- QIAGEN GmbH Coordinator
- Medical University of Graz (Prof. K. Zatloukal)
- University of Florence (Prof. M. Pazzagli)
- CIRMMP Florence, CERM (Prof. I. Bertini)
- TATAA Biocenter
- PreAnalytiX GmbH
- DIAGENIC ASA
- Aros Applied Biotechnology
- Dako Denmark
- ACIES
- Biotechnology Inst. of Czech Academy of Science (Prof. M. Kubista)
- European Committee for Standardization (CEN)
- ImmunID Technologies
- Erasmus Medical Center Rotterdam (Prof. P. Riegman)
- Technical University Munich (*Prof. H. Hoefler, Prof. K. Becker*)
- Fondazione IRCCS Istituto Nazionale dei Tumori (Dr. P. Verderio)

Scientific Advisory Board

- Prof. François Rousseau (Univ. Laval, Quebec. CanGeneTest Network)
- Dr. Roberta M. Madej (CLSI)

Project Ethics Committee

- Dr. Anne Cambon-Thomsen (CNRS, INSERM, Tolouse, France)
- Dr. Ruth Chadwick (ESRC Centre, Cardiff University, UK)



SPIDIA Consortium Bi-Annual Meeting Berlin November 2011





Thank you!

Questions?

