

Lab ID: LXXX

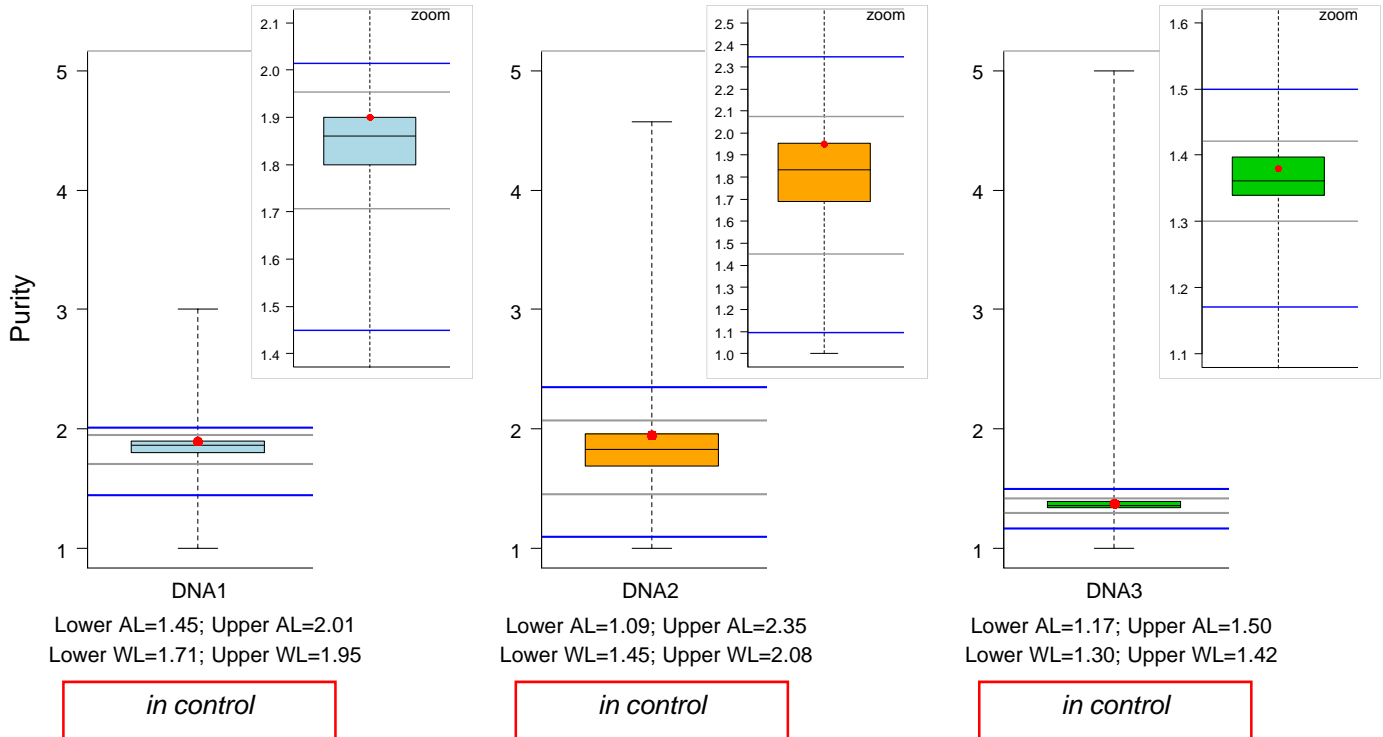
A. Purity and concentration of DNA1, DNA2 and DNA3 (pre-extracted DNAs)

A.1 Spectrophotometric data provided by your lab

| | 260nm | 280nm | 320nm | Purity | Concentration (ng/μl) | Dilution factor |
|------|-------|-------|-------|--------|-----------------------|-----------------|
| DNA1 | 1.343 | 0.705 | . | 1.90 | 67.14 | 1 |
| DNA2 | 0.357 | 0.183 | . | 1.95 | 17.85 | 1 |
| DNA3 | 1.574 | 1.137 | . | 1.38 | 78.72 | 1 |

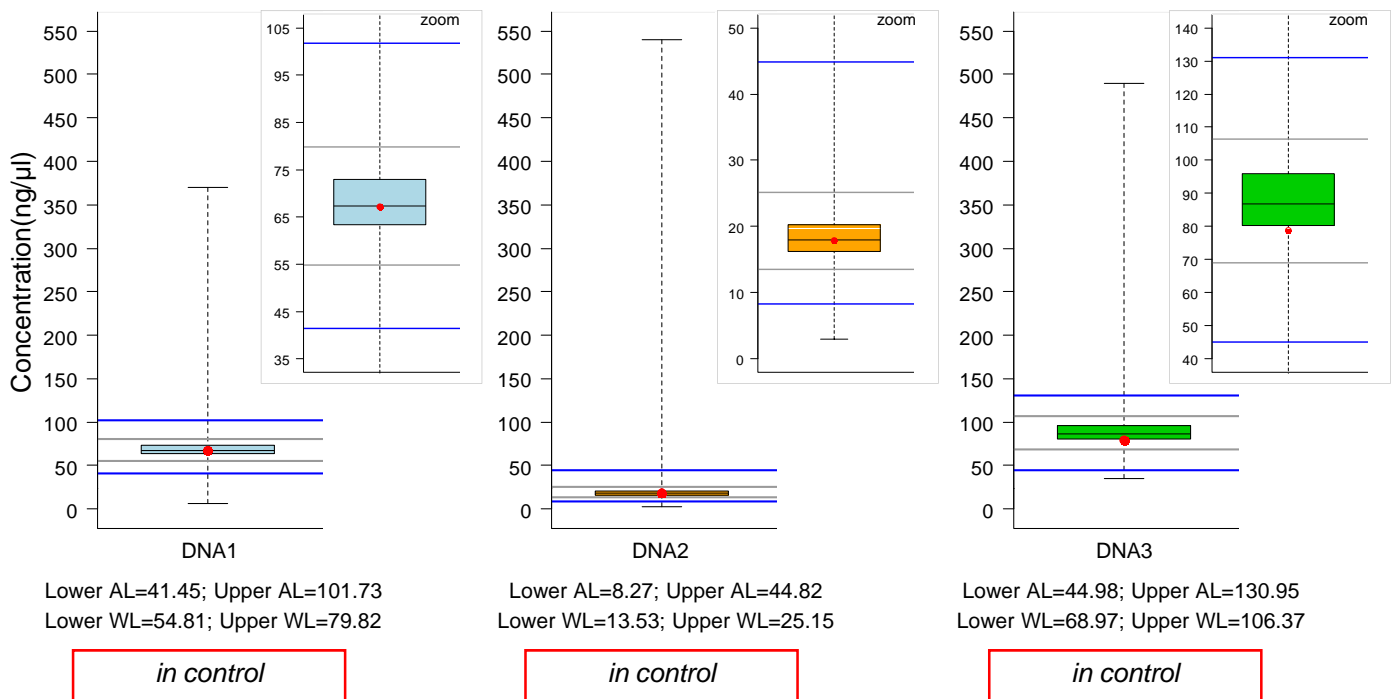
A.2 Your lab (●) versus overall distribution (N=172) – Purity

In the figures the blue lines represent the Action Limits (ALs) and the gray lines represent the Warning Limits (WLs).



A.3 Your lab (●) vs overall distribution (N=174) – Concentration

In the figures the blue lines represent the Action Limits (ALs) and the gray lines represent the Warning Limits (WLs).



Lab ID: LXXX

B. Purity and Quantity of DNA4 (DNA extracted from blood)

B.1 Spectrophotometric data provided by your lab and by SPIDIA lab

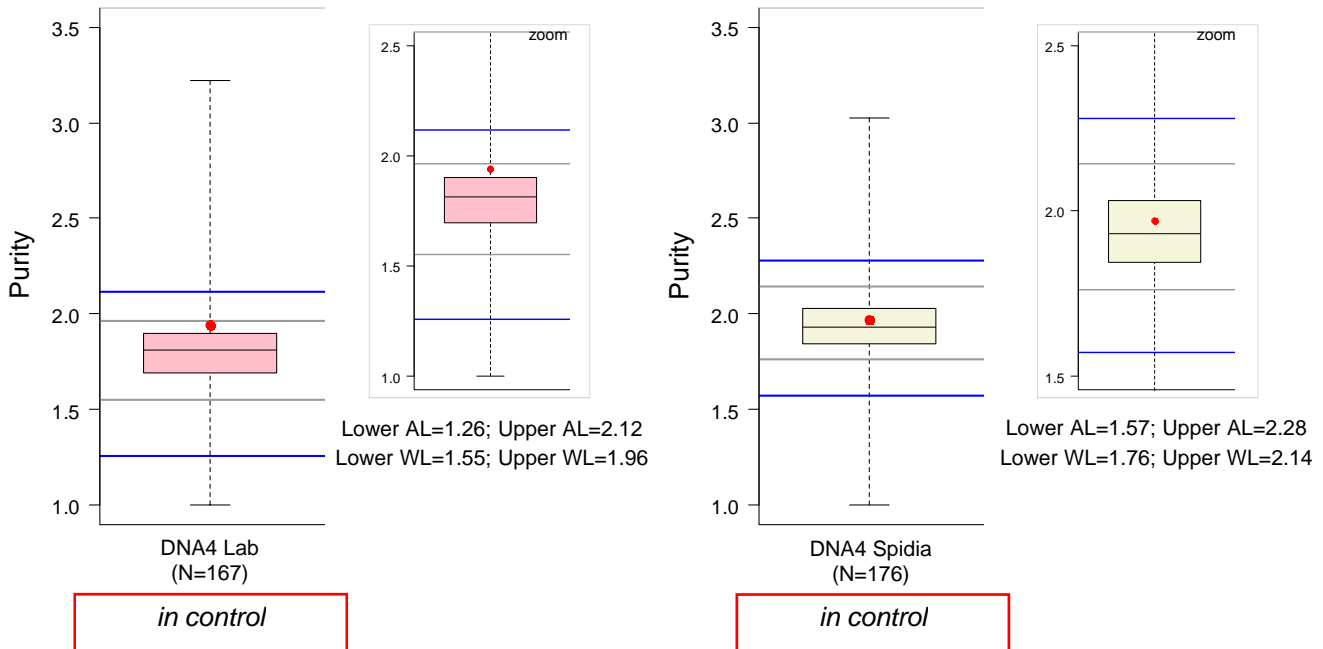
| 260nm Lab | 280nm Lab | 320nm Lab | Purity Lab | Quantity (ng/μl) Lab | Purity Spidia | Quantity (ng/μl) Spidia | Dilution factor | Extraction vol. (ul) | Elution vol. (ul) | Buffer |
|-----------|-----------|-----------|------------|----------------------|---------------|-------------------------|-----------------|----------------------|-------------------|---------|
| 2.868 | 1.482 | . | 1.94 | 21.507 | 1.970 | 26.858 | 1 | 1000 | 150 | TE 10.1 |

B.2 Additional information provided by your lab

| Extraction | | Spectrophotometer | | Temperature of DNA storage | | Time interval (hours) | |
|------------|----------|-------------------|------------------|----------------------------|------------------------|-----------------------|------------------------|
| producer | supplier | producer | supplier | arrival to extraction | extraction to analysis | arrival to extraction | extraction to analysis |
| Homebrew | X | Labtech | Nanodrop ND-1000 | -20 °C | 20 °C | 19 h | 72 h |

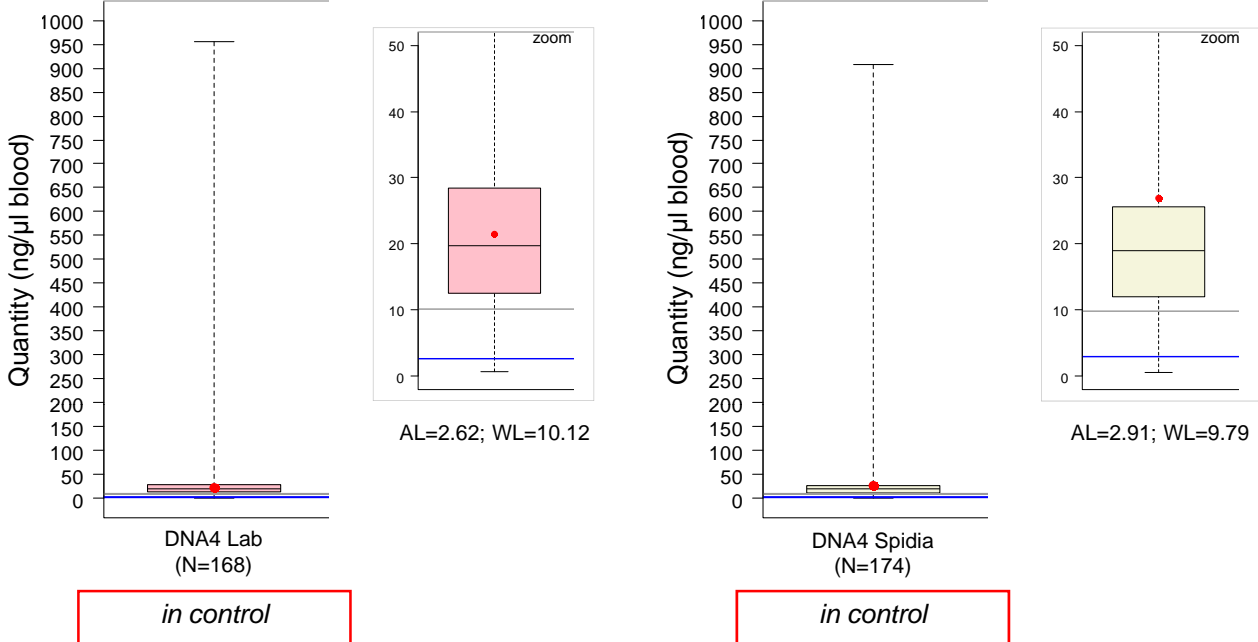
B.3 Your lab (●) versus overall distribution – Purity

In the figures the blue lines represent the Action Limits (ALs) and the gray lines represent the Warning Limits (WLs).



B.4 Your lab (●) versus overall distribution - Quantity

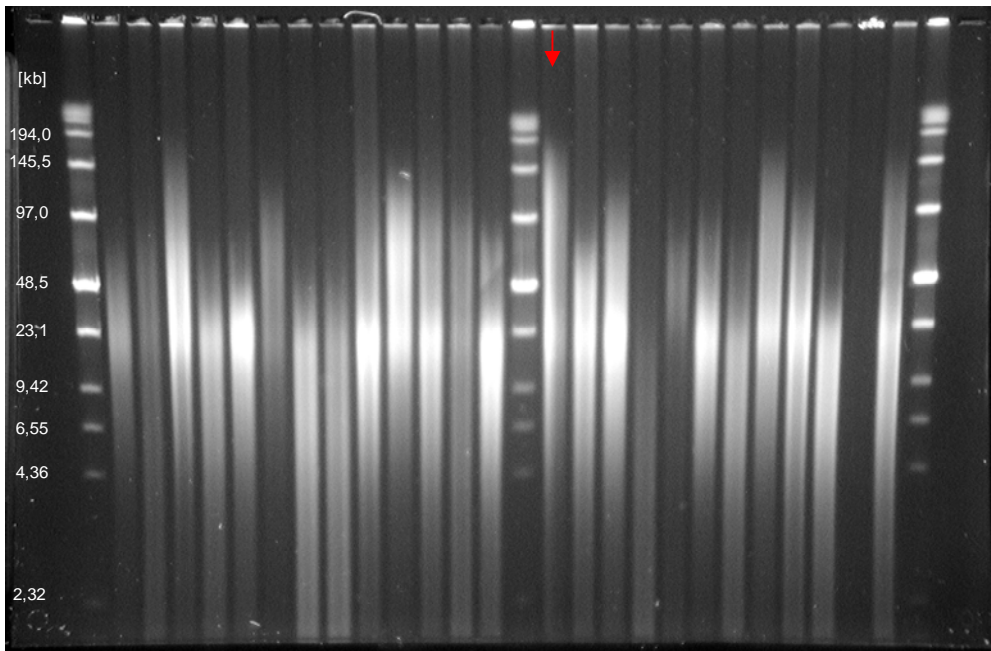
In the figures the blue line represents the Action Limit (AL) and the gray line represents the Warning Limit (WL).



Lab ID: LXXX

C. Integrity of DNA4 (DNA extracted from blood)

C.1 Your lab (↓) - Pulse field gel electrophoresis image

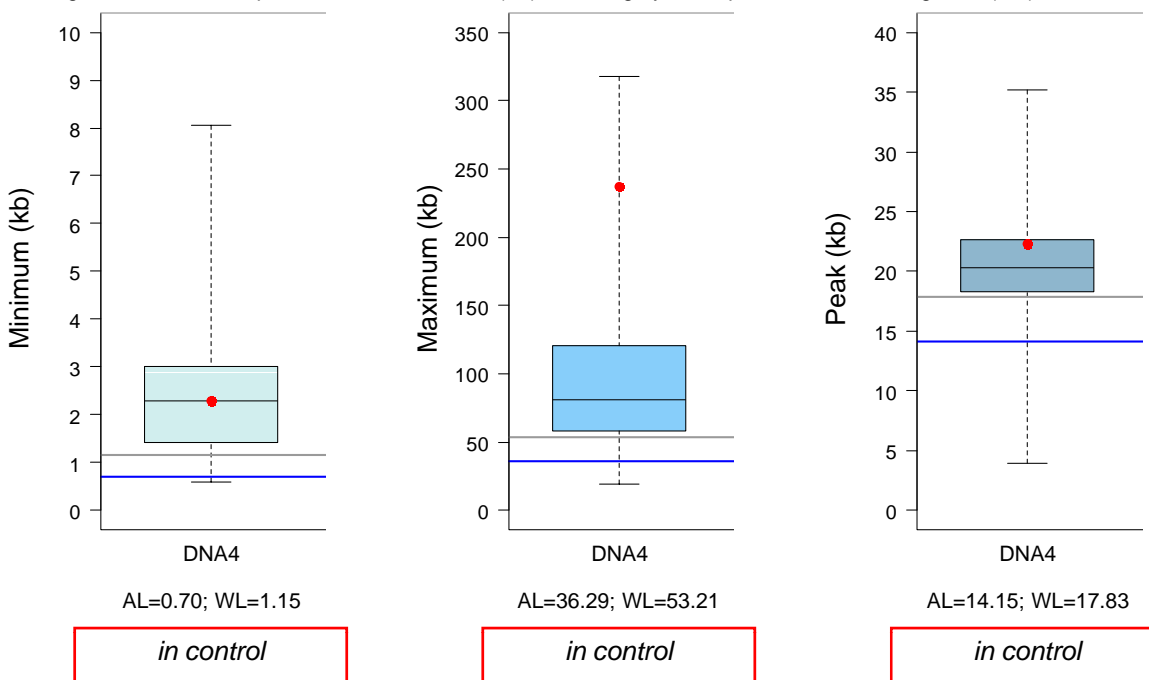


C.2 ImageJ data of your lab

| Minimum (Kb) | Maximum (Kb) | Peak (Kb) |
|--------------|--------------|-----------|
| 2.299 | 236.800 | 22.317 |

C.3 Your lab (●) versus overall distribution (N=157) – ImageJ data

In the figure the blue line represents the Action Limit (AL) and the gray line represents the Warning Limit (WL).



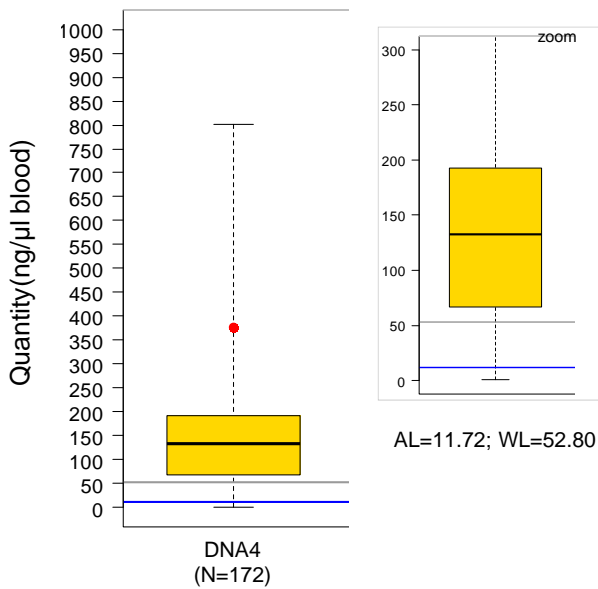
Lab ID: LXXX

D. Quantification of RNaseP by real-time PCR on DNA4 and evaluation of interferences

D.1 Your lab (●) versus overall distribution

Quantification of RNaseP

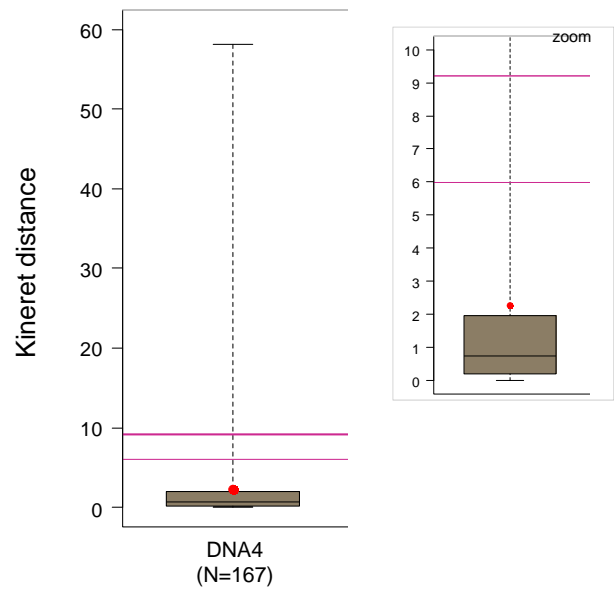
In the figure the blue line represents the Action Limit (AL) and the gray line represents the Warning Limit (WL).



in control

Interferences

In the figure the two lines represent the two Kineret threshold for outliers identification: 5.99 (weak outlier) and 9.21 (strong outlier).



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Lab ID: LXXX

E. Integrity of DNA5 (DNA extracted from plasma)

E.1 Isohelix/Agilent analysis - Ratio integrity

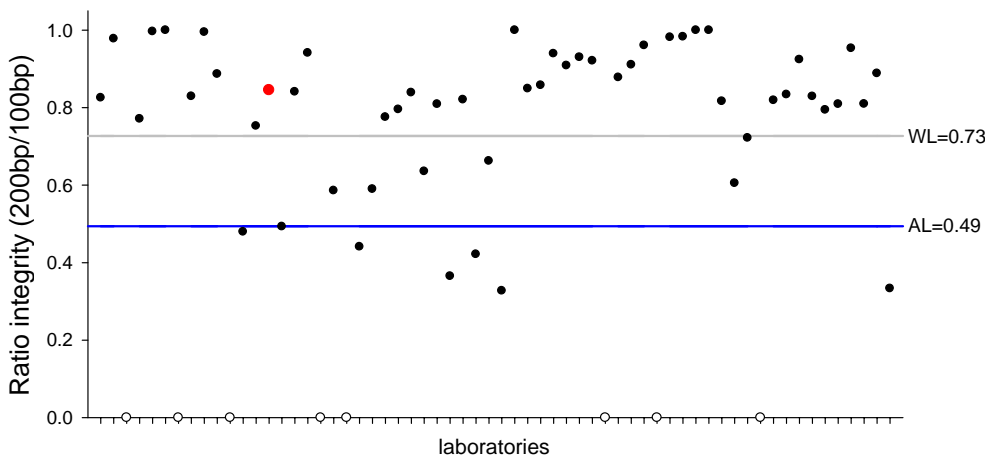
| Ratio integrity 200bp/100bp | Ratio integrity 300bp/100bp | Ratio integrity 400bp/100bp | Extraction vol. (ul) | Elution vol. (ul) | Buffer |
|-----------------------------|-----------------------------|-----------------------------|----------------------|-------------------|---------|
| 0.845 | 0.647 | 0.526 | 1000 | 50 | TE 10.1 |

E.2 Additional information provided by your lab

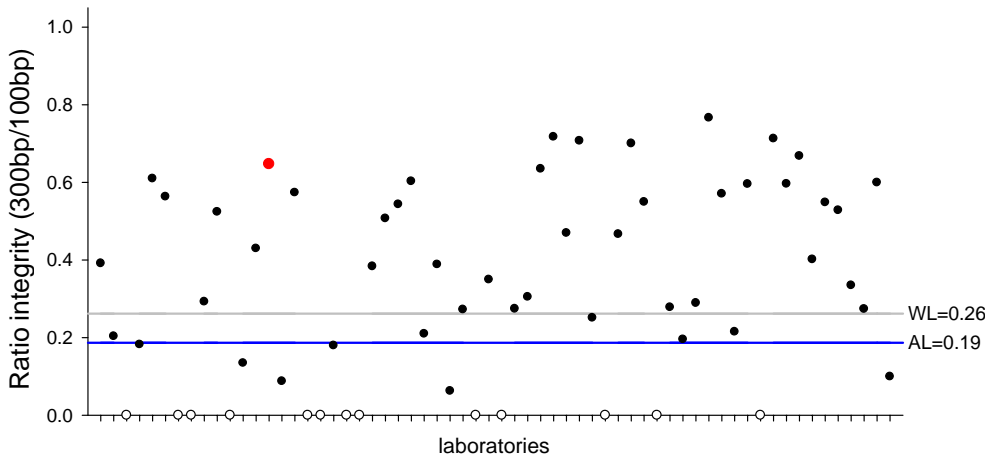
| Extraction | | Temperature of DNA storage | | Time interval (hours) | |
|--------------------------|--------------|----------------------------|------------------------|-----------------------|------------------------|
| producer | supplier | arrival to extraction | extraction to analysis | arrival to extraction | extraction to analysis |
| QIAmp DNA Blood mini kit | QIAGEN 51106 | -20 °C | 20 °C | 19 h | 72 h |

E.3 Your lab (●) versus overall distribution (N=62) – Ratio integrity

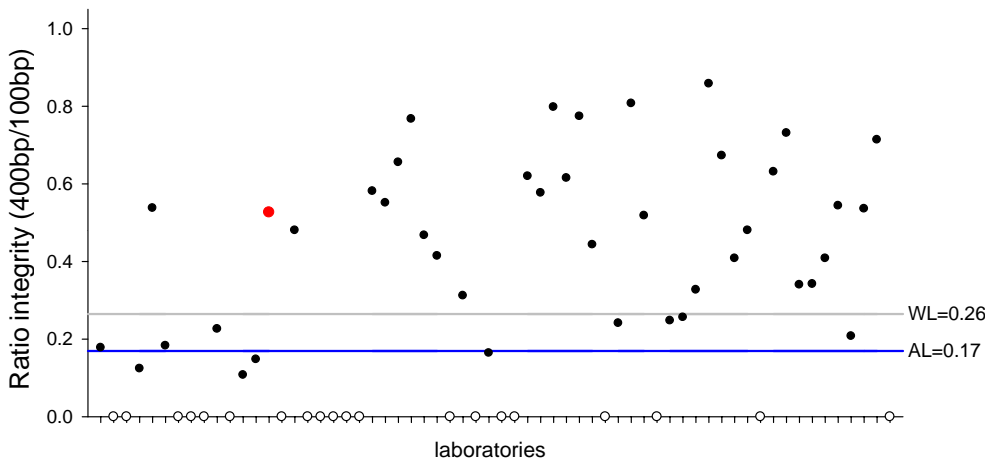
In the figures the blue line represents the Action Limit (AL) and the gray line represents the Warning Limit (WL).



in control



in control



in control

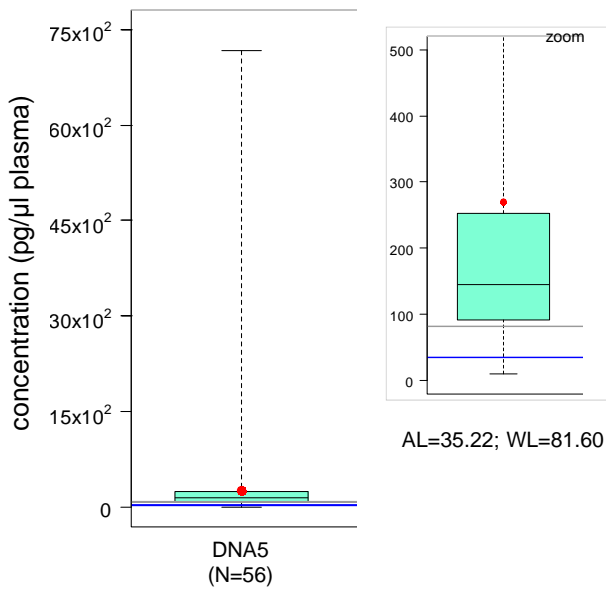
Lab ID: LXXX

F. Quantification of RNaseP by real-time PCR on DNA5 and evaluation of interferences

F.1 Your lab (●) versus overall distribution

Quantification of RNaseP

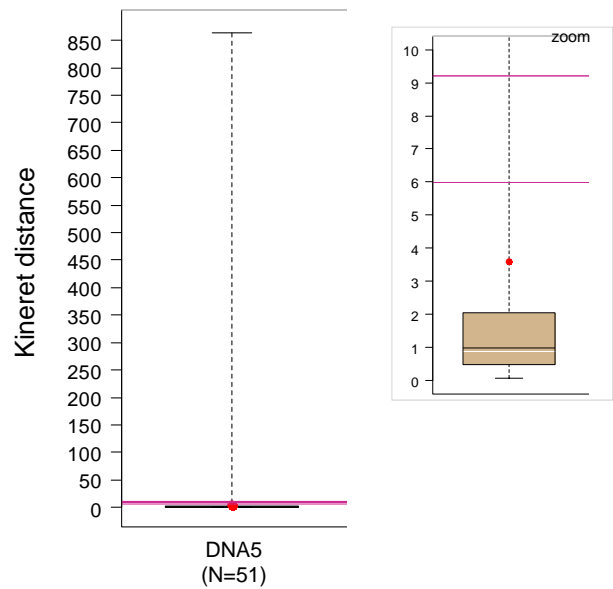
In the figure the blue line represents the Action Limit (AL) and the gray line represents the Warning Limit (WL).



in control

Interferences

In the figure the two lines represent the two Kineret threshold for outliers identification: 5.99 (weak outlier) and 9.21 (strong outlier).

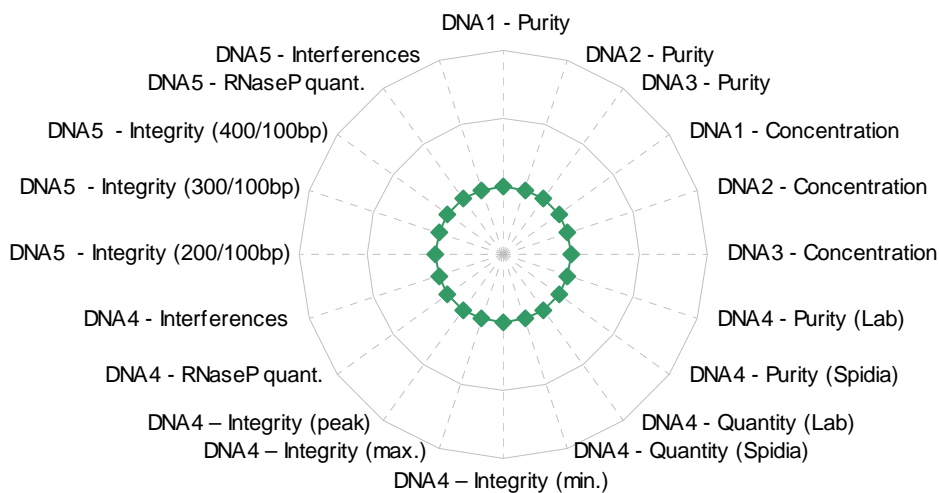


in control

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G. Summary

| | Performance | | Missing | Comments |
|------------------------------|-------------|--|---------|----------|
| DNA1 – Purity | in control | | | |
| DNA2 – Purity | in control | | | |
| DNA3 – Purity | in control | | | |
| DNA1 – Concentration | in control | | | |
| DNA2 – Concentration | in control | | | |
| DNA3 – Concentration | in control | | | |
| DNA4 – Purity (Lab) | in control | | | |
| DNA4 – Purity (Spidia) | in control | | | |
| DNA4 – Quantity (Lab) | in control | | | |
| DNA4 – Quantity (Spidia) | in control | | | |
| DNA4 – Integrity (min.) | in control | | | |
| DNA4 – Integrity (max.) | in control | | | |
| DNA4 – Integrity (peak) | in control | | | |
| DNA4 – RNaseP quant. | in control | | | |
| DNA4 – Interferences | in control | | | |
| DNA5 – Integrity (200/100bp) | in control | | | |
| DNA5 – Integrity (300/100bp) | in control | | | |
| DNA5 – Integrity (400/100bp) | in control | | | |
| DNA5 – RNaseP quant. | in control | | | |
| DNA5 – Interferences | in control | | | |



This report has been produced with the collaboration of the **University of Florence** (M.Pazzagli, S.Gelmini, C.Orlando, L.Simi, F.Malentacchi), **Fondazione IRCCS Istituto Nazionale dei Tumori of Milan** (P.Verderio, S.Pizzamiglio, C.Ciniselli), **QIAGEN** (R.Wyrich, C.Hartmann) and **TATAA BIOCENTER** (A.Tichopad).